KAMIYA BIOMEDICAL COMPANY

ACE Inhibition Screening Kit

For the measurement of ACE inhibitory activity

Cat. No. KT-534

For Research Use Only. Not for Use in Diagnostic Procedures.

PRODUCT INFORMATION

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PRODUCT
The **K-*ASSAY*®** ACE Inhibition Screening Kit is for the measurement of ACE inhibitory activity.

BACKGROUND
Angiotensin-converting enzyme (ACE) is one of the key elements responsible for vasopressor action. ACE converts angiotensin I to angiotensin II, a potent vasopressor, in the renin-angiotensin system and contributes to increasing blood pressure by inactivating bradykinin, a strong antihypertensive peptide. Recently, various functional foods have received attention because of their inhibitory activity toward ACE.

ACE activity is conventionally determined by UV measurement of the hippuric acid produced from the synthetic substrate Hyppuryl–His–Leu. However, the assay process is complicated and requires organic solvent. In this kit, a safe and straightforward modified method has been developed.

The colorimetric detection system in the kit determines the amount of 3-hydroxybutyric acid (3HB) generated from 3-hydroxybutyryl–Gly–Gly–Gly by ACE. The kit is designed for 96-well microplate assays and is suitable for multiple sample measurements. No organic solvent extraction is required. The assay is safe, simple, and provides highly reproducible data.

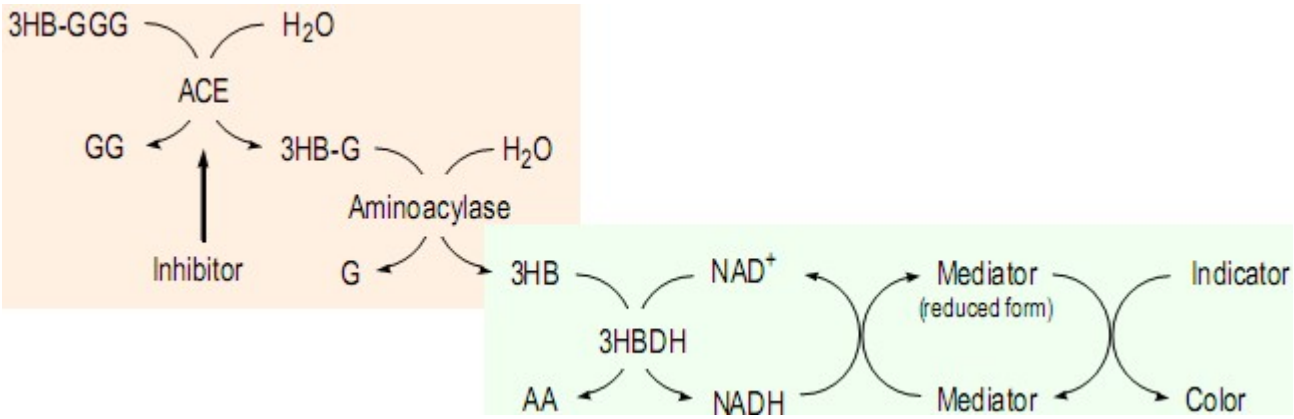


Figure 1. Principle of ACE Inhibitory activity assay using the **K-*ASSAY*®** ACE Inhibition Screening Kit

COMPONENTS
- Substrate Buffer (1 mL), 2 vials
- Enzyme A, 2 vials
- Enzyme B, 2 vials
- Enzyme C, 2 vials
- Coenzyme, 2 vials
- Indicator solution (5 mL), 2 vials

MATERIALS OR EQUIPMENT REQUIRED BUT NOT PROVIDED
- Microplate reader (450 nm filter).
- 96-well microplate.
- 2 - 20 µL, 20 - 200 µL & 100 - 1,000 µL pipettes
- Multi-channel pipette
- Incubator
- Disposable syringes (1 mL)

PREPARATION OF WORKING SOLUTION

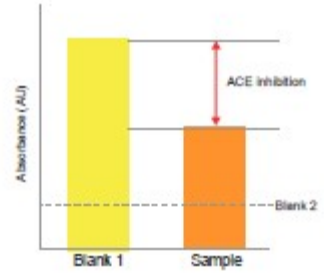
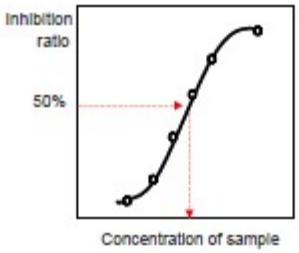
A. Enzyme working solution

Dissolve Enzyme B in 2 mL of deionized water to prepare Enzyme B solution. Then add 1.5 mL of Enzyme B solution to Enzyme A to prepare Enzyme working solution.
Note: Enzyme A and B vials are capped under vacuum pressure. Add deionized water or solution through the rubber septum with a syringe, and then remove the septum.
Note: The Enzyme working solution is stable at −20°C for 2 weeks or in a refrigerator for 3 days.

B. Indicator working solution

Dissolve Enzyme C and Coenzyme in 3 mL of deionized water each. Add 2.8 mL of Enzyme C solution and 2.8 mL of Coenzyme solution to Indicator solution to prepare Indicator working solution.
Note: Enzyme C and Coenzyme vials are capped under vacuum pressure. Add deionized water through the rubber septum with a syringe, and then remove the septum.
Note: The Indicator working solution is stable at −20°C for 2 weeks or in a refrigerator for 3 days.

SELECTION OF PROTOCOL

Protocol 1	*Protocol 2*
Confirmation of the presence or absence of ACE inhibition	Determination of IC_{50}
	
28 samples can be measured in triplicate.	4 samples can be measured in triplicate.

PROTOCOL 1

Confirmation of the presence or absence of ACE inhibition

Sample Preparation
Prepare 100 µL of each sample.
Note: If the sample volume is <100 µL, please dilute the sample.
Note: If the sample is colored, prepare 150 µL of each sample.

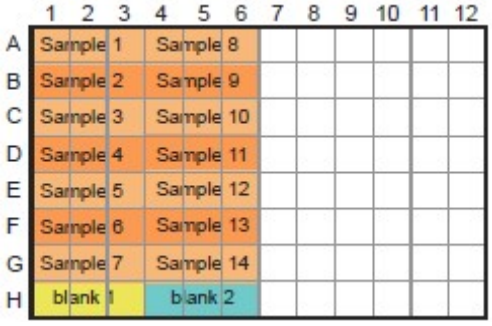
Procedure for Colorless Sample
See Table 1 and Figure 2
1) Add 20 µL of sample solution to each sample well.
2) Add 20 µL of deionized water to each blank 1 well and 40 µL to each blank 2 well.
3) Add 20 µL of Substrate buffer to each well.
4) Add 20 µL of Enzyme working solution to each sample well and blank 1 well.
Note:
Enzyme working solution is easy to remain on the well wall. Please tap the plate to ensure that all solutions have been mixed completely.
Since the enzymatic reaction starts immediately after adding the Enzyme working solution, use a multi-channel pipette to minimize the well-to-well time lag.
5) Incubate at 37°C for 1 h.
6) Add 200 µL of Indicator working solution to each well.
7) Incubate at room temperature for 10 min.
8) Read the absorbance at 450 nm using a microplate reader.

Table 1 Amount of sample and reagent needed for each well.

	Sample	blank 1	blank 2
Sample solution	20 µl	-	-
Deionized water	-	20 µl	40 µl
Substrate buffer	20 µl	20 µl	20 µl
Enzyme working solution	20 µl	20 µl	-
Indicator working solution	200 µl	200 µl	200 µl

blank 1: positive control (no ACE inhibition)
blank 2: reagent blank

Figure 2 Example of arrangement on a 96-well microplate.



Confirmation of the presence or absence of ACE inhibition
ACE inhibition can be calculated from the equation:

ACE inhibition (%) = [(Ablank 1 − Asample)/(Ablank 1 − Ablank 2)] × 100

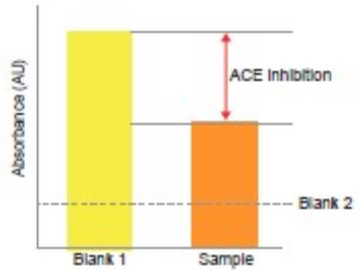


Figure 3 Confirmation of the presence or absence of ACE inhibition.

Procedure for Colored Sample
See Table 2 and Figure 4
1) Add 20 µL of sample solution to each sample well and sample blank well.
2) Add 20 µL of deionized water to each blank 1 well, 40 µL to each blank 2 well and 240 µL to each sample blank well.
3) Add 20 µL of Substrate buffer to each sample well, blank 1 well and blank 2 well.
4) Add 20 µL of Enzyme working solution to each sample well and blank 1 well.

Note:
Enzyme working solution is easy to remain on the well wall. Please tap the plate to ensure that all solutions have been mixed completely.
Since the enzymatic reaction starts immediately after adding the Enzyme working solution, use a multi-channel pipette to minimize the well-to-well time lag.
5) Incubate at 37°C for 1 h.
6) Add 200 µL of Indicator working solution to each sample well, blank 1 well and blank 2 well.
7) Incubate at room temperature for 10 min.
8) Read the absorbance at 450 nm using a microplate reader.

Table 2 Amount of sample and reagent needed for each well.

	Sample	blank 1	blank 2	Sample blank
Sample solution	20 µl	-	-	20 µl
Deionized water	-	20 µl	40 µl	240 µl
Substrate buffer	20 µl	20 µl	20 µl	-
Enzyme working solution	20 µl	20 µl	-	-
Indicator working solution	200 µl	200 µl	200 µl	-

blank 1: positive control (no ACE inhibition)
blank 2: reagent blank

Figure 4 Example of arrangement on a 96-well microplate.

	1	2	3	4	5	6	7	8	9	10	11	12
A	Sample 1			Sample 8			Sample blank 1			Sample blank 8		
B	Sample 2			Sample 9			Sample blank 2			Sample blank 9		
C	Sample 3			Sample 10			Sample blank 3			Sample blank 10		
D	Sample 4			Sample 11			Sample blank 4			Sample blank 11		
E	Sample 5			Sample 12			Sample blank 5			Sample blank 12		
F	Sample 6			Sample 13			Sample blank 6			Sample blank 13		
G	Sample 7			Sample 14			Sample blank 7			Sample blank 14		
H	blank 1			blank 2								

Confirmation of the presence or absence of ACE inhibition
ACE inhibition can be calculated from the equation:
ACE inhibition (%) = $[(A_{blank\ 1} - A_{sample} - A_{sample\ blank})/(A_{blank\ 1} - A_{blank\ 2})] \times 100$

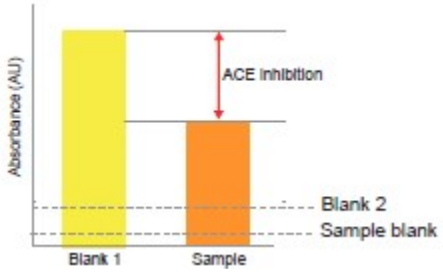


Figure 5 Confirmation of the presence or absence of ACE inhibition

PROTOCOL 2

Determination of IC_{50}

Preparation of Sample solution

Dilute sample solution with deionized water.
Dilutions: 1 (no dilution), 1/5, $1/5^2$, $1/5^3$, $1/5^4$. $1/5^5$, $1/5^6$

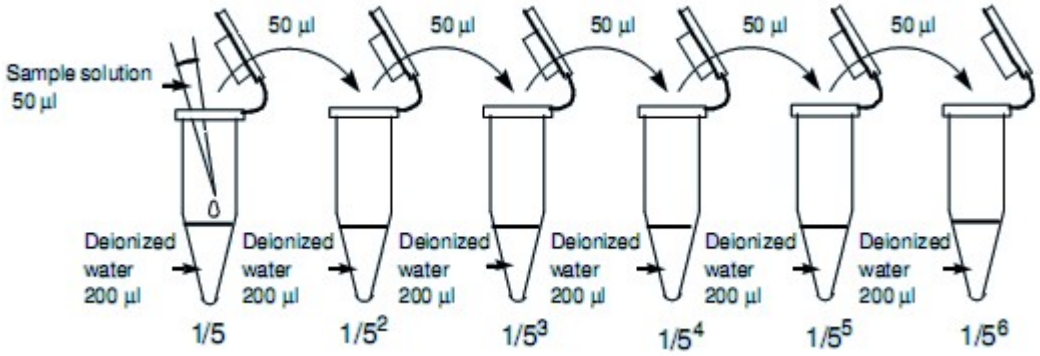


Figure 6. Preparation of sample solutions

Procedure for Colorless Sample

See Table 3 and Figure 7

1) Add 20 µL of sample solution to each sample well.
2) Add 20 µL of deionized water to each blank 1 well and 40 µL to each blank 2 well.
3) Add 20 µL of Substrate buffer to each well.
4) Add 20 µL of Enzyme working solution to each sample well and blank 1 well.

Note:

Enzyme working solution is easy to remain on the well wall. Please tap the plate to ensure that all solutions have been mixed completely.

Since the enzymatic reaction starts immediately after the addition of the Enzyme working solution, use a multi-channel pipette to minimize the well-to-well time lag.

5) Incubate at 37°C for 1 h.
6) Add 200 µL of Indicator working solution to each well.
7) Incubate at room temperature for 10 min.
8) Read the absorbance at 450 nm using a microplate reader.
9) ACE inhibition can be calculated from the equation:

ACE inhibition (%) = [(Ablank 1 − Asample)/(Ablank 1 − Ablank 2)] × 100

Table 3 Amount of sample and reagent needed for each well.

	Sample	blank 1	blank 2
Sample solution	20 µl	-	-
Deionized water	-	20 µl	40 µl
Substrate buffer	20 µl	20 µl	20 µl
Enzyme working solution	20 µl	20 µl	-
Indicator working solution	200 µl	200 µl	200 µl

blank 1: positive control (no ACE inhibition)
blank 2: reagent blank

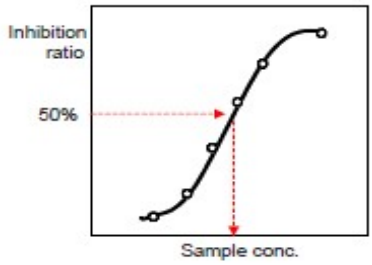
Figure 7 Example of arrangement on a 96-well microplate.

	1	2	3	4	5	6	7	8	9	10	11	12
A	Sample		1	Sample		2						
B	Sample		1/5	Sample		2/5						
C	Sample		$1/5^2$	Sample		$2/5^2$						
D	Sample		$1/5^3$	Sample		$2/5^3$						
E	Sample		$1/5^4$	Sample		$2/5^4$						
F	Sample		$1/5^5$	Sample		$2/5^5$						
G	Sample		$1/5^6$	Sample		$2/5^6$						
H	blank		1	blank		2						

Determination of IC50 (50% inhibitory concentration)

- Prepare an inhibition curve using sample concentration for the *x*-axis and percentage inhibition of ACE for the *y*-axis. A typical inhibition curve is shown in Figure 8.
- Determine the concentration of the sample solution that gives 50% ACE inhibition as indicated in Figure 8.
- Because the total volume of the inhibition assay is 60 µL (first step of the assay), the original sample is diluted 3 times in the reaction. Therefore, the actual concentration of the sample at 50% inhibition is one-third of the concentration determined from the inhibition curve.

Figure 8 Inhibition curve



Procedure for Colored Sample

See Table 4 and Figure 9

1) Add 20 µL of sample solution to each sample well and sample blank well.
2) Add 20 µL of deionized water to each blank 1 well, 40 µL to each blank 2 well and 240 µL to each sample blank well.
3) Add 20 µL of Substrate buffer to each sample well, blank 1 well and blank 2 well.
4) Add 20 µL of Enzyme working solution to each sample well and blank 1 well.

Note:

Enzyme working solution is easy to remain on the well wall. Please tap the plate to ensure that all solutions have been mixed completely.

Since the enzymatic reaction starts immediately after adding the Enzyme working solution, use a multi-channel pipette to minimize the well-to-well time lag.

5) Incubate at 37°C for 1 h.
6) Add 200 µL of Indicator working solution to each sample well, blank 1 well and blank 2 well.
7) Incubate at room temperature for 10 min.
8) Read the absorbance at 450 nm using a microplate reader.
9) ACE inhibition can be calculated from the equation.

ACE inhibition (%) = [(Ablank 1 − Asample − Asample blank) / (Ablank 1 − Ablank 2)] × 100

Table 4 Amount of sample and reagent needed for each well.

	Sample	blank 1	blank 2	Sample blank
Sample solution	20 µl	-	-	20 µl
Deionized water	-	20 µl	40 µl	240 µl
Substrate buffer	20 µl	20 µl	20 µl	-
Enzyme working solution	20 µl	20 µl	-	-
Indicator working solution	200 µl	200 µl	200 µl	-

blank 1: positive control (no ACE inhibition)
blank 2: reagent blank

Figure 9 Example of arrangement on a 96-well microplate

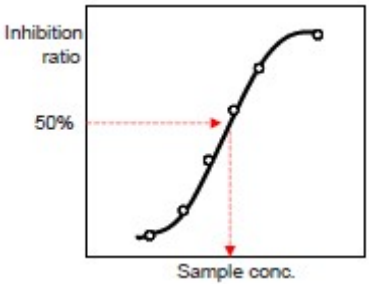
	1	2	3	4	5	6	7	8	9	10	11	12
A	Sample		1	Sample		2	Sample blank 1			Sample blank 2		
B	Sample		1/5	Sample		2/5	Sample blank 1/5			Sample blank 2/5		
C	Sample		$1/5^2$	Sample		$2/5^2$	Sample blank $1/5^2$			Sample blank $2/5^2$		
D	Sample		$1/5^3$	Sample		$2/5^3$	Sample blank $1/5^3$			Sample blank $2/5^3$		
E	Sample		$1/5^4$	Sample		$2/5^4$	Sample blank $1/5^4$			Sample blank $2/5^4$		
F	Sample		$1/5^5$	Sample		$2/5^5$	Sample blank $1/5^5$			Sample blank $2/5^5$		
G	Sample		$1/5^6$	Sample		$2/5^6$	Sample blank $1/5^6$			Sample blank $2/5^6$		
H	blank 1			blank 2								

Determination of IC50 (50% inhibitory concentration)

• Prepare an inhibition curve using sample concentration for the *x*-axis and percentage inhibition of ACE for the *y*-axis. A typical inhibition curve is shown in Figure 10.

• Determine the concentration of the sample solution that gives 50% ACE inhibition as indicated in Figure 10.
• Because the total volume of the inhibition assay is 60 µL (first step of the assay), the original sample is diluted 3 times in the reaction. Therefore, the actual concentration of the sample at 50% inhibition is one-third of the concentration determined from the inhibition curve.

Figure 10 Inhibition curve



STORAGE

Store the kit at 4°C.

PRECAUTION

- Several kit components are in glass vials. Please handle with care.
- Multiple measurements (triplicates of each sample) are recommended to obtain accurate data.
- If the water solubility of the sample is low, use dimethylsulfoxide or ethanol to dissolve. Then, dilute the solution with an appropriate buffer. The final concentration of organic solvent should be <1%.
- If the sample solution is acidic, adjust the pH to ≥5 before use for measurement.
- Ascorbic acid may interfere with the assay. The concentration of ascorbic acid in the sample solution should be <0.01% w/v. If the sample solution contains insoluble materials, remove it by centrifugation or filtration before use for measurement.

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