



# KAMIYA BIOMEDICAL COMPANY

# Osteoclast Culture Kit

For the culture of Osteoclasts from precursor cells.

Cat. No.: KT-646 KT-647

For Research Use Only.



### PRODUCT INFORMATION

## Osteoclast Culture Kit Cat. No. KT-646 KT-647

#### **PRINCIPLE**

In aging societies, osteoporosis and other age-related bone metabolism disorders are a rapidly increasing problem. The amount of bone in an organism is controlled by a balance of osteoblasts (bone-forming cell) and osteoclasts (bone-destroying cell) activities. A method to induce osteoclasts formation from bone marrow cells using M-CSF (macrophage-colony stimulating factor) and RANKL (receptor activator of NF-kB ligand) has been established in recent years. This kit includes cryopreserved primary precursor osteoclasts from mouse bone marrow and Culture Medium containing M-CSF and RANKL.

#### **COMPONENTS**

Components	KT-646	KT-647
Mouse Osteoclast Precursor Cells, frozen	2 vials containing 2 x 10 <sup>6</sup> cells each	1 vial containing 2 x 10 <sup>6</sup> cells
Washing Medium	50 mL	50 mL
Culture Medium, M-CSF (10 ng/mL) and RANK Ligand (10 ng/mL)	25 mL	25 mL
Osteoplate	2 plates	2 plates

### Materials required but not provided

- Pipettes
- Tubes
- Refrigerated centrifuge
- Water bath
- Von Kossa or toluidine blue stains for pit image analysis

#### **PRECAUTIONS**

- 1. Read the instructions carefully before beginning the culture.
- 2. This kit is for research use only, not for human or diagnostic use.

Primary precursor osteoclasts are shipped on dry ice. If not used immediately, store in liquid nitrogen.

#### **PROTOCOL**

- 1. Thaw the Wash and Culture Medium in a 37°C water bath with gentle shaking.
- 2. Quickly thaw a vial of primary precursor osteoclasts in a 37°C water bath.
- 3. Transfer thawed cells to a 15 mL centrifuge tube, add 10 mL of Wash Medium and mix gently.
- 4. Centrifuge at 1000 rpm (170xg) for 5 minutes at 4°C.
- 5. Remove supernatant and add 10 mL of Wash Medium and mix gently.
- 6. Centrifuge at 1000 rpm (170xg) for 5 minutes at 4°C.
- 7. Remove supernatant and resuspend the cells in 2.5 5 mL of Culture Medium. To study factors that effect osteoclasts formation, add the factors to the Culture Medium.
  - a. If the cells are resuspended in 5 mL of Culture Medium, there will be enough cell suspension for about 50 wells.
  - b. To quickly observe osteoclasts formation, culture the cells at a higher density.
- 8. Transfer 100  $\mu$ L of cell suspension into each well of a 96-well plate.
- 9. Incubate at 37°C, 5% CO2, 100% humidity.

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- 10. Replace the medium with fresh culture medium every 3-4 days. Cells will begin to fuse and form osteoclasts after 4 days of incubation (fig 1).
- 11. Count the osteoclasts by staining with tartrate-resistant acid phosphatase (TRAP Staining Kit, Cat. No. KT-008) or Pit image analysis.

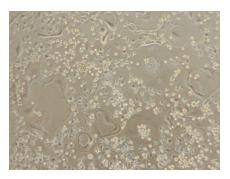
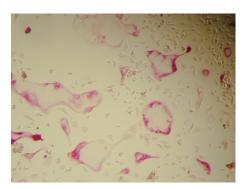


Figure 1: Osteoclasts differentiation

#### **EXAMPLES**

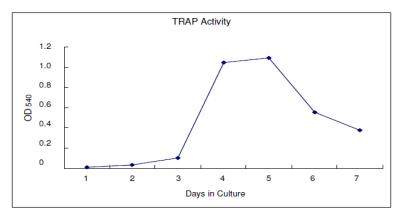
TRAP Staining Kit (Cat. No. KT-008):
 Osteoclasts were fixed then stained with 5 mL of a mixture containing chromogenic substrate and tartrate-containing buffer.



**TRAP Staining** 

2. Quantitation of TRAP in culture supernatant (Cat. No. KT-008):

Thirty microliters of culture supernatant was incubated for 3 hours in the presence of chromogenic substrate/tartrate-containing buffer. The samples were read at wavelength 540 nm.



Measurement of TRAP in Osteoclasts culture supernatant

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#### 3. Pit Image Analysis

Pits should be observed prior to 10 days in culture and before the cells dissolve the osteoplate coating.

- A. Aspirate the medium completely from wells. Add 100 uL/well of a 10% bleach solution.
- B. Incubate in the bleach solution for 5 minutes at room temperature.
- C. Aspirate bleach solution and wash each well twice with 150 uL of dH2O.
- D. Allow the plate to air dry at room temperature (Recommended time 3 to 5 hours).
- E. Observe each well at 100x magnification for the formation of pits.
- F. Pits will appear as individual or multiple clusters at the bottom of the well.
- G. Analyze data appropriately; recommended methods include:
  - -Visual enumeration of pits via a microscope or analysis software.
  - -Stain using Von Kossa or toluidine blue stains to increase contrast between pits and surface coating.

#### 4. Scanning electron microscopy (SEM):



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