



KAMIYA BIOMEDICAL COMPANY

Chicken Anti-Sheep Red Blood Cell (SRBC) IgG ELISA

For the quantitative determination of SRBC IgG in chicken serum and plasma

Cat. No. KT-576

For research use only, not for use in diagnostic procedures.

PRODUCT INFORMATION**Chicken Anti-Sheep Red Blood Cell (SRBC) IgG ELISA**
Cat. No. KT-576**PRODUCT**

The **K-ASSAY®** Chicken SRBC IgG ELISA is for the quantitative determination of SRBC IgG in chicken serum or plasma.

PRINCIPLE

The Chicken Anti-SRBC IgG ELISA is based on a solid phase enzyme-linked immunosorbent assay (ELISA). The assay uses detergent solubilized SRBC ghosts for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated anti-chicken IgG antibodies for detection. Test serum or plasma samples are diluted and incubated in the microtiter wells for 45 minutes. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 45 minutes. Anti-SRBC IgG molecules are thus sandwiched between immobilized SRBC antigens and the detection antibody conjugate. The wells are then washed to remove unbound HRP-labeled antibodies and TMB reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of stop solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of anti-SRBC IgG is proportional to the optical density of the test sample.

COMPONENTS

- Microtiter Plate: SRBC coated 96-well plate (12 strips of 8 wells)
- Enzyme Conjugate Solution: 11 mL
- Calibrator: Lyoph.
- Diluent Buffer: 50 mL
- TMB Solution: 11 mL
- Stop Solution: 11 mL, 1N HCl
- Wash Buffer (20x): 50 mL

Materials or Equipment required but not provided

- Plate reader (450 nm)
- Micropipette and tips
- De-ionized water
- Graph paper (PC software is optional)
- Paper towels
- Polypropylene or glass tubes
- Vortex mixer
- Plate shaker/incubator
- Plate washer

STORAGE

Store at 4°C. Microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air. The kit is stable until the expiration date when stored as noted in this section.

General Instructions

1. Please read and understand the instructions thoroughly before using the kit.
2. This kit is designed to measure anti-SRBC IgG levels in chicken serum or plasma collected 5-7 days after immunization with SRBC.
3. All reagents should be allowed to reach room temperature (18-25°C) before use.
4. The optimal sample dilution should be determined empirically. However, studies suggest an initial sample dilution of 500 fold.
5. Serum and plasma samples must be diluted at least 15 fold in diluent.
6. Optimum results are achieved if, at each step, reagents are pipetted into wells of the microtiter plate within 5 minutes.

PREPARATION OF REAGENTS

Wash Buffer

The wash solution is provided as 20x stock. Prior to use dilute the contents of the bottle (50 mL) with 950 mL of distilled or deionized water.

Calibrator

1. Reconstitute the vial of lyophilized chicken anti-SRBC IgG calibrator stock with 200 μL of diluent and then prepare the 100 u/mL calibrator in a polypropylene or glass tube as described on the vial label.
2. Label 4 polypropylene or glass tubes as 50, 25, 12.5, and 6.25 u/mL, and pipette 250 μL of diluent into each tube.
3. Prepare a 50 u/mL calibrator by diluting and mixing 250 μL of the 100 u/mL calibrator with 250 μL of diluent in the tube labelled 50 u/mL.
4. Similarly prepare the 25, 12.5, and 6.25 u/mL calibrators by serial dilution.
5. Note: After reconstitution the calibrator stock solution is stable for at least one week if stored at 4°C. It should be frozen at or below -20°C if use beyond this time is intended. Avoid multiple freeze-thaws.

SAMPLE PREPARATION

Note: Studies indicate that anti-SRBC IgG is present in chicken serum or plasma at concentrations of approximately 10,000 u/mL seven days after immunization with SRBC. In order to obtain values within range of the calibration curve, we suggest samples initially be diluted 500 fold using the following procedure for each sample tested. Optimal dilutions may need to be determined empirically.

1. For each test sample dispense 90 μL and 245 μL of diluent into separate tubes.
2. Pipette and mix 10 μL of the serum/plasma sample into the tube containing 90 μL of diluent. This provides a 10 fold diluted sample.
3. Mix 5 μL of the 10 fold diluted sample with 245 μL diluent in the second tube to give a 500 fold dilution of sample.
4. Repeat this procedure for each sample to be tested.

PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μL of calibrators, and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μL /well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μL of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
8. Wash as detailed in 4 and 5 above.
9. Dispense 100 μL of TMB reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature for 20 minutes.
11. Stop the reaction by adding 100 μL of Stop Solution to each well.
12. Gently mix. It is important to make sure all the blue color changes to yellow.
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

CALCULATION OF RESULTS

1. Calculate the average absorbance values for each set of calibrators, and samples.
2. Construct a calibration curve by plotting the mean absorbance obtained from each calibrator against its concentration in u/mL on linear graph paper, with absorbance values on the vertical or Y axis and concentrations on the horizontal or X axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of anti-SRBC IgG in u/mL from the calibration curve.
4. Multiply the derived concentrations by the dilution factor to determine the actual concentration for anti-SRBC IgG in the serum/plasma sample.
5. PC graphing software may be used for the above steps.
6. If the OD values of samples fall outside the calibration curve when tested at a dilution of 100, samples should be diluted appropriately and re-tested.

Limitations of the Procedure

- Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
- The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

FOR RESEARCH USE ONLY**KAMIYA BIOMEDICAL COMPANY**

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