

KAMIYA BIOMEDICAL COMPANY

Human esRAGE ELISA

**For the quantitative determination of human esRAGE in serum, plasma and
tissue culture medium**

Cat. No. KT-1882

For Research Use Only. Not for use in diagnostic procedures.

PRODUCT INFORMATION**Human esRAGE ELISA****Cat. No. KT-1882****INTENDED USE**

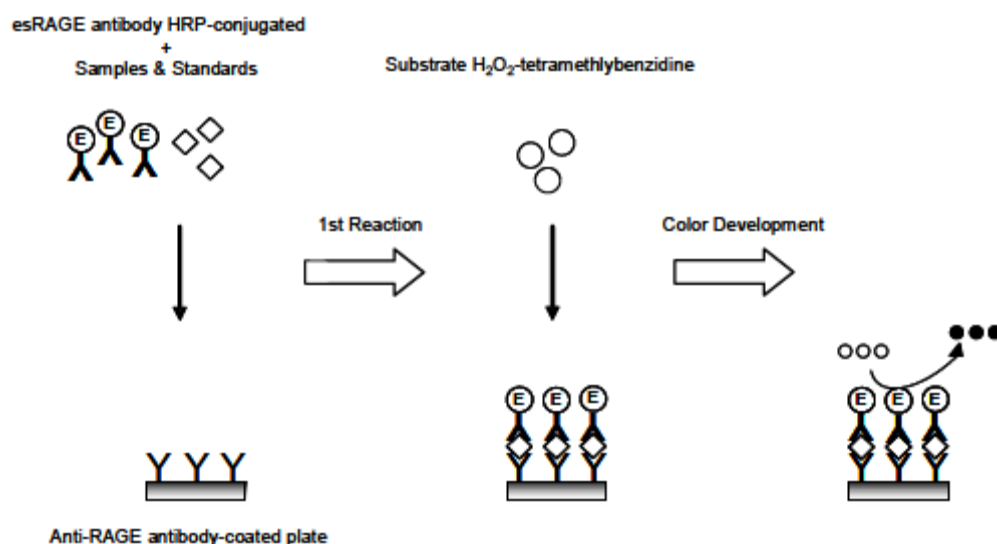
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INTRODUCTION

In multicellular organisms, most cellular and plasma proteins undergo a nonenzymatic glycation by the covalent binding of reduced sugars to the amino groups on proteins. A balance exists between early glycation products and non-glycated proteins during homeostasis. Under the condition of prolonged hyperglycemia as in diabetes, these early stage glycation products undergo additional chemical rearrangements to form irreversibly bound advanced glycation end products (AGE). AGE has been shown to be a major factor in vascular cell derangement in diabetic samples. AGE has also been implicated in Alzheimer's Disease by the glycation of amyloid- β peptide and microtubule-associated protein tau. The receptor for AGE (RAGE), a 55 kDa protein and a member of the immunoglobulin superfamily of cell surface molecules, is found in mononuclear phagocytes, endothelium, vascular smooth muscle, and the central nervous system. RAGE is a multi-ligand receptor. The most pathological consequence of the AGE-RAGE engagement is induction of oxidative stress, activation of nuclear factor κ B (NF- κ B), resulting in disruption of homeostatic functions in the vasculature. AGE has also been shown to up-regulate the RAGE expression in endothelial cells.

Translation of RAGE mRNA isolated from human endothelial cells revealed a novel splice variant named endogenous secretory receptor (esRAGE) which can capture AGE and neutralize the effects of AGE on cells. Endogenous secretory RAGE has been found circulating in blood and in extracellular fluids of our bodies.

The principle of the assay is shown in Figure 1. Calibrators or samples and detection antibody (esRAGE antibody horseradish peroxidase conjugated) are incubated in an anti-RAGE antibody coated 96-well microtiter plate. After 16-24 hour incubation and four washing, substrate is added to each well. The plate is incubated for 30 minutes. The enzymatic reaction is stopped by the addition of stop solution and absorbance of the resulting yellow product is measured at 450 nm. The absorbance is proportional to the concentration of esRAGE. A calibration curve is constructed by plotting absorbance values versus esRAGE concentrations of calibrators, and concentrations of unknown samples are determined using this calibration curve.

Figure 1. Assay Principle

COMPONENTS**1. PRIMARY ANTIBODY-COATED PLATE** 1 Plate

One 96-well plate with adsorbed anti-RAGE antibody
Plate is provided in a resealable foil pouch with desiccant.

2. DETECTION ANTIBODY 1 Bottle (11 mL)

esRAGE antibody-horseradish peroxidase (HRP) conjugated

3. SUBSTRATE 1 Bottle (15 mL)

3,3',5,5'-tetramethylbenzidine and hydrogen peroxide

4. STOP SOLUTION 1 Bottle (15 mL)

1 M sulfuric acid

5. BUFFER 1 Bottle (5 mL)

Phosphate buffered saline containing bovine serum albumin

6. esRAGE CALIBRATOR 1 Bottle (lyophilized)

Human esRAGE, 1.6 ng

7. 5X WASH SOLUTION 2 Bottle (50 mL each)

Phosphate buffered saline with Tween 20

MATERIALS REQUIRED BUT NOT PROVIDED

- Graduated cylinder
- Micropipette(s) and disposable pipette tips
- Null strips for 96-well plate
- 96-well plate or manual strip washer
- Paper towels or absorbent paper
- Plate reader capable of measuring absorbance at a wavelength of 450 nm (reference filter at 650 nm, optional)
- Well-closed containers such as microtubes (1.5 mL or more in capacity)
- Refrigerator, 4°C

REAGENT PREPARATION

Allow all the reagents to come to room temperature (20-30 °C) prior to the start of reagent preparation.

1. 1X Wash Solution

If crystals are observed in the 5X Wash Solution, warm the bottle in a 37°C water bath until the crystals disappear. Prepare 1X Wash Solution by mixing 50 mL of the 5X Wash Solution with 200 mL of de-ionized water or equivalent. Mix well. After preparation, store 1X Wash Solution at 4°C. The 1X Wash Solution is stable for 1 month at 4°C.

2. esRAGE Calibrator

Reconstitute esRAGE calibrator with 0.5 mL of distilled water. Prepare each esRAGE assay calibrator (1.6 ng/mL, 0.8 ng/mL, 0.4 ng/mL, 0.2 ng/mL, 0.1 ng/mL, 0.05 ng/mL) by serially diluting the reconstituted calibrator with buffer. Use the undiluted esRAGE Calibrator (3.2 ng/mL) and buffer for the 3.2 ng/mL and 0 ng/mL assay calibrators, respectively.

Assay calibrators should be prepared prior to use and any unused assay calibrators should be discarded.

Reconstituted esRAGE calibrator is stable for 1 month at 4°C.

3. Other kit reagents

Anti-RAGE antibody-coated plate, esRAGE antibody horseradish peroxidase (HRP)-conjugated, substrate, stop solution, and buffer are ready for use. Once opened, these reagents are stable for 3 months at 4°C.

Note: The unopened reagents, if properly maintained at 4°C, are stable until the expiration date printed on the box label.

Do not mix reagents from different kits unless they have the same lot number.

SAMPLE PREPARATION

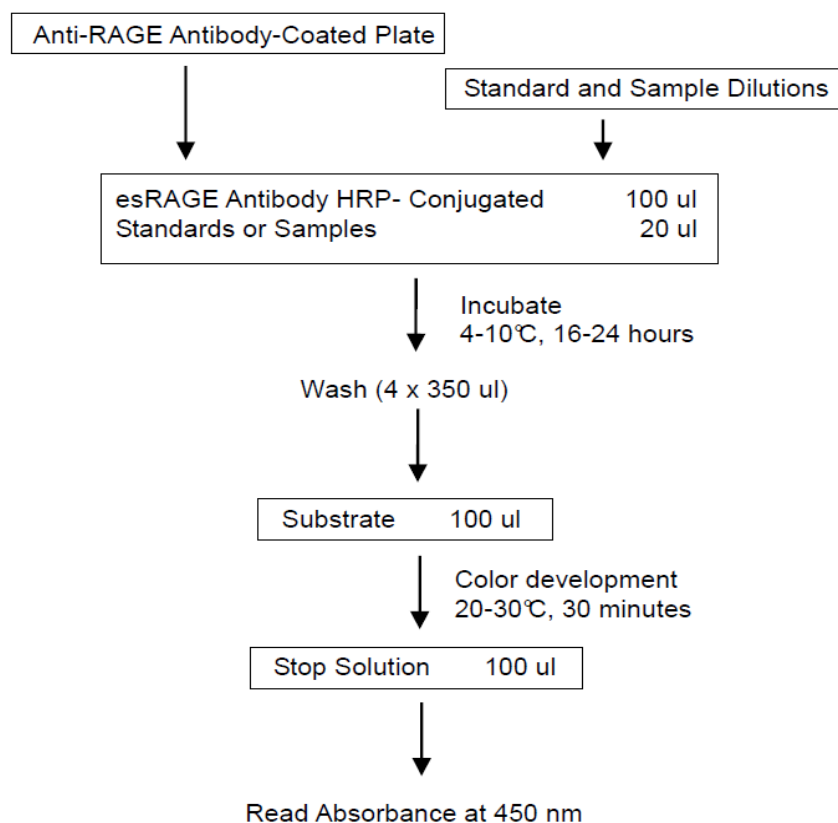
Samples may be diluted with buffer, if quantitation of the sample protein is above the 3.2 ng/mL data point on the calibration curve.

ASSAY PROTOCOL

Note: Allow all reagents to come to room temperature (20-30°C) prior to the start of the assay and prepare 1X Wash Solution and esRAGE Calibrators as described in the previous sections.

1. Remove Primary Antibody-Coated Plate from its foil pouch. Remove any unneeded strips from the plate frame, reseal them in the foil pouch, and return the foil pouch to 4 °C. If a 96-well plate washer is used, the plate frame should be completely filled with wells by adding as many null strips as necessary. Identify well position(s) for each sample on a data sheet or plate map.
2. Add 100 µL of esRAGE antibody HRP-conjugate to the appropriate number of antibody-coated wells.
3. Add 20 µL of assay calibrators (0, 0.05, 0.1, 0.2, 0.4, 0.8, 1.6, 3.2 ng/mL) or sample to the plate and mix well by pipetting. All assay calibrators and samples should be run in duplicates. Every plate must include the assay calibrators to properly correlate the sample readings.
4. Cover plate(s) securely with a plate sealer and incubate at 4-10°C for 16-24 hours. The plates should not be agitated or rocked during the incubation period.
5. Aspirate the reaction mixture from each well.
6. Wash each well with 350 µL of 1x wash solution.
7. Aspirate the wash solution
8. Repeat steps 6 and 7 three times for a total of 4 washes
9. Invert the plate and gently tap on a clean absorbent towel to remove residual droplets of wash solution.
10. Dispense 100 µL of substrate into each well.
11. Avoid exposing the microtiter plate to direct sunlight. Covering the plate with aluminum foil is recommended. Incubate at 20-30°C for 30 minutes.
12. Add 100 µL of stop solution to each well.
13. Read the plate within 30 minutes after stopping the reaction. The plates are measured at wavelength 450 nm. The recommended reference wavelength is 630 nm.

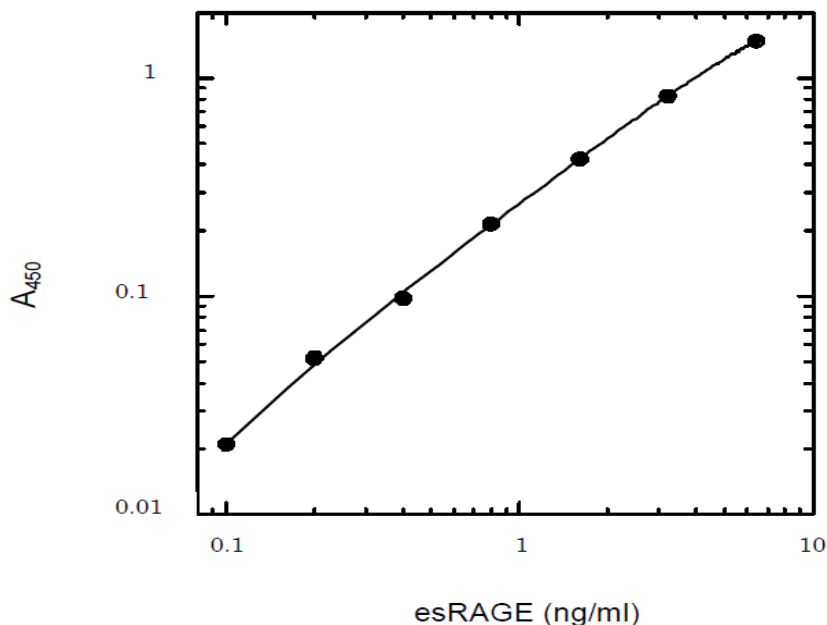
Figure 2. Flow Chart of Assay



RESULTS

1. Subtract the mean absorbance value of the 0 ng/mL blank from each mean absorbance value of the calibrator series and samples tested (Net Absorbance).
2. Plot the log of known concentrations of each calibrator and the calculated Net Absorbance on the X-axis and Y-axis, respectively. Fit an appropriate regression curve to the plotted points.
3. Determine the esRAGE concentrations of the samples by interpolation of the regression curve formula.
4. If samples were diluted, then concentration calculations must be multiplied by the appropriate dilution factor to obtain the correct results for the undiluted samples.

Figure 3. Typical Calibration Curve



STORAGE

Store all components at 4°C. DO NOT FREEZE.

TROUBLESHOOTING

1. Lack of signal or weak signal in all wells

Possible explanations:

- Omission of a reagent or a step.
- Improper preparation or storage of a reagent.
- Assay performed before reagents were allowed to come to 20-30°C.
- Plate reader did not perform well.

2. High signal and background in all wells

Possible explanations:

- Improper or inadequate washing; be certain that all wash volumes and repetitions were correct.
- Improper dilution of detection antibody.
- Overdeveloping; decrease the incubation time before the Stop Solution is added.

3. High background in sample wells only

Possible explanations:

- Sample concentration was too high.
- Improper dilution of detection antibody.

4. Weak signal in sample wells only

Possible explanations:

- Sample concentration was too low.
- Improper dilution of detection antibody.

FOR RESEARCH USE ONLY

KAMIYA BIOMEDICAL COMPANY

12779 Gateway Drive, Seattle, WA 98168

Tel: (206) 575-8068 Fax: (206) 575-8094

Email: LifeScience@k-assay.com

www.k-assay.com